

# Antimicrobial activity of black seed oil & water extracts On multidrug resistant *Pseudomonas aeruginosa*

Muna S. Al-Delaimi

University of Duhok - Faculty of Medical Sciences.



## ARTICLE INFO

Received: 20 / 6 /2012  
Accepted: 26 / 8 /2012  
Available online: 29/8/2013  
DOI: [10.37652/juaps.2012.78250](https://doi.org/10.37652/juaps.2012.78250)

### Keywords:

Burn infection,  
MDRPa,  
acetone extract,  
black seed oil.

## ABSTRACT

*Pseudomonas aeruginosa* is most common cause of nosocomial infections in burn centers. This opportunistic and multidrug resistant bacterium causes severe problems for hospitalized burn patients. Samples were collected from patients attending Burn & Plastic Surgery Hospital in Duhok / Iraq, between September 2011 and March 2012, for preliminary identification of *P. aeruginosa* using conventional methods, then confirmed with the use of BD -Phoenix™ Automated Microbiology System. Isolates were considered Multi Drug Resistant *Pseudomonas aeruginosa* (MDRPa) if they showed resistance to the three or more classes of antipseudomonal agents. Accordingly thirty isolates were found MDRPa. The pattern of resistance revealed that the highest resistance was for Gentamicin 96.7%, Cefepime 96.7%, Ceftazidime 90% and Aztreonam 83.3%, while lowest resistance was detected against Amikacin 36.7% and Colistin 3.3%. The results of disc diffusion assay demonstrated that antibacterial activities of black seed oil acetone extract at 1:1,1:10,1:25 concentrations were found effective against all tested MDRPa isolates, which were statistically significant ( $P < 0.05$ ), while at highest dilution 1:75. All MDRPa isolates showed lowest activity of oil extract. The antimicrobial effects of water extract showed lowest and poor activity against all tested bacteria.

## Introduction:

Burn injury is a major public health problem throughout the world.<sup>1</sup> These injuries still produce significant morbidity and mortality in developing countries.<sup>2</sup> Infection in burn patients is difficult to control due to the presence of dead and denatured burn eschar, and moist environment, that act as a good growth medium for microbes.<sup>3</sup> Due to prolonged hospital stay these patients are at high risk of nosocomial infection. In this situation topical antimicrobial agents play a limited role that reduces the incidence of septic complication but the incidence of bacterial colonization had not decreased.<sup>4,5</sup> *Pseudomonas aeruginosa* is one of the most commonly isolated pathogens, and is the most frequently isolated non fermentative bacillus in clinical specimens.<sup>6</sup> It is also a part of normal skin flora of humans but can cause life threatening opportunistic infections specially in immunocompromised hosts.

This organism is a significant cause of burn and nosocomial infections. The ability of *Pseudomonas aeruginosa* to destroy tissue may be related to the production of various extracellular enzymes.<sup>7</sup> Complicating the empiric selection of adequate therapy is the increasing prevalence of antimicrobial drug resistance among *P. aeruginosa*, it is estimated that as many as 75% of all deaths following burn injury are related to infection.<sup>8</sup>

An alarming increase in bacterial strains resistant to existing antimicrobial agents demands a renewed effort to seek agents effective against pathogenic bacteria resistant to current antimicrobials.<sup>9</sup> The seeds of *Nigella sativa* L. (Ranunculaceae), commonly known as black seed or black cumin, are used in folk (herbal) medicine all over the world for the treatment and prevention of a number of diseases. Its seeds have a great medicinal importance and have been reported to exhibit many pharmacological effects that include anti-parasitic, antibacterial, antifungal, antiviral, antioxidant and anti-inflammatory activities.<sup>10</sup>

\* Corresponding author at: University of Duhok - Faculty of Medical Sciences. E-mail address:

However only a limited data is available for its efficacy against multi drug resistant *P. aeruginosa*, which isolated from different clinical specimens.<sup>11,12,13</sup>

The present study was therefore designed to determine the antimicrobial activity of different concentrations of black seed extracts against MDRPa isolated from burn patients, it was the first trial in our country

## Materials & methods:

### Isolation, characterisation and identification

The study was performed at Burn & Plastic Surgery Hospital in Duhok / Iraq, between September 2011 and March 2012. Burned patients were enrolled in this study and from each one a cotton swabs collected to preliminary identification of *Pseudomonas aeruginosa* using colony structure and colony morphology, gram stain, oxidase positive reaction, typical smell, and development of pyocyanin pigments.<sup>14</sup> Then confirmed with the use of BD - Phoenix™ Automated Microbiology System (BD Diagnostics System, Sparks, MD, USA) / Bacteriological Laboratory / Azadi teaching Hospital, which is designed for the rapid identification (ID)(Table 1) and quantitative determination of antimicrobial susceptibility testing (AST)( Table 2) by Minimal Inhibitory Concentration (MIC) of Gram Negative aerobic and facultative anaerobic bacteria from pure culture.<sup>15</sup> Isolates were considered Multi Drug Resistant *Pseudomonas aeruginosa* (MDRPa) if they showed resistant to the three or more classes of antipseudomonal agents.<sup>16,17,18</sup> Accordingly thirty isolates were found as MDRPa .

### Preparation of black seed extracts

Plant seed were purchased from a local market in the city of Duhok. The extraction was conducted at Chemistry Department\ Faculty of Science –University of Duhok. To obtain the oil extract, the seed were powdered mechanically using as blender for five minutes. A total of 80 grams of the powder was dissolved in 100 ml cold acetone; the mixture was then filtered using Whatman filter paper number 41. The filtrate was left to dry in an open Petridish at room temperature for 24 hours to allow the solvent to evaporate, then transferred into sterile vials (1ml of each) and stored at -200C till used. The following concentration (1:1, 1:10, 1:25, 1:50, 1:75) of oil

extract were obtained by diluting the pure oil in absolute alcohol.

Black seed water extract, was done by the modified method of Ibraheem. 19 : 80 gm powder of black seed was placed in sterile beaker containing 100ml of distilled water. A magnetic stirrer was used to continuously mix the suspension for 18 hours. The mixture was then filtered using Whatman filter paper No. 41. The filtrate was left to dry in an open dish for 24 hours , and 0.5 g was dissolved in 100 ml distilled water to obtain a stock solution of 0.5% w/v of the extract. Concentrations of 0.05% w/v, 0.025% w/v and 0.005% w/v were prepared from the stock solution by adding distilled water.

### Preparation of the drug impregnated filter paper disc

Whatman Filter paper No. 1 was used to prepare discs (6 mm). The discs were then sterilized by autoclaving. During sensitivity testing, 4µl of both oil extract in pure or diluted form and aqueous extract was kept on filter paper disc, placed on Mueller Hinton Agar plate inoculated with bacteria (Fig.3). Prepared discs were stored at 4°C in the refrigerator till used. Disc containing absolute alcohol (diluent of the extract) was used as negative control.

### Inoculation of plates:

This was done by the modified method of Pelczar *et al*<sup>20</sup> using flood-inoculation technique. Bacterial suspension in Nutrient Broth having turbidity equivalent 0.5 McFarland was freshly prepared and 2 ml of this was transferred onto the Mueller Hinton (MH) Agar plate and distributed gently over the surface of medium with gentle rocking. The excess fluid was removed from the plate and the plate was kept in incubator at 37°C for 30 minutes for drying before application of discs. A fully sensitive strain of *Pseudomonas aeruginosa* was used as standard for the black seed oil and water extract discs of anti *Pseudomonas* susceptibility test.

### Antimicrobial susceptibility testing

The antibiotic susceptibility pattern of the *Pseudomonas aeruginosa* isolates was determined using the disk diffusion method according to the modified Kirby-Bauer technique, by placing discs impregnated with test material on surface of inoculated Muller Hinton agar plates. The plates were then kept in incubator at 37°C for 18 hours and diameters of

zones of inhibition were measured. The experiment was performed in triplicate.

Statistical Analysis:

Done with SPSS ver.18 for ANOVA one way.

### Results:

During the study period thirty MDRPa were isolated which showed different resistance to various antibacterial drugs as follows : four were resistant to four antibiotics, eight to five antibiotics, five to six antibiotics , four to eight antibiotics, four to nine antibiotics and five to ten antibiotics (Table 1).

The overall pattern of resistance revealed that the highest resistance was for Gentamicin 96.7% (29/30), Cefepime 96.7% (29/30), Ceftazidime 90% (27/30) and Aztreonam 83.3% ( 25/30) .Ciprofloxacin 63.3%(19/30), Levofloxacin 60% (18/30) , Piperacillin 60%(18/30), while lowest resistance was detected against Imipenem 46.7% (14/30) , Meropenem 43.3% (13/30), Amikacin 36.7%(11/30) and Colistin 3.3% (1/30) ( Fig. 4).

The present investigation revealed that the acetone extract yield 20-30% of black seed oil, is being reported for the first time, it was reddish brown liquid , oily in nature.

Antibacterial activities of black seed oil extract at it's different concentrations against a total of thirty MDRPa strains were shown in (Table-2) .Five different concentrations 1:1, 1:10 , 1:125 ,1:50 and 1:75 per disc were used , among them 1:1,1:10 and 1:125 were found more effective against all isolates (mean of diameter of inhibition zone were  $23.08 \pm 1.401\text{mm}$  ,  $19.482 \pm 2.014\text{ mm}$  and  $14.553 \pm 3.421\text{ mm}$  respectively ) which was found statistically significant ( $P < 0.05$ ).

Table-3 revealed that the oil extract inhibited MDRPa isolates in dose dependent manner. In conclusion this table indicate that the three concentrations (1:1, 1:10,1:25) of oil extract had resulted in highest antibacterial activity (16 - >25mm) and remaining 7 strains showed lowest activity pattern ( $\leq 10\text{mm}$ ) at dilution 1:25. Out of thirty resistant isolates tested with dilution (1:50) showed activity against 9 isolates (11-15mm), and the remaining 21 isolates showed lowest activity pattern ( $\leq 10\text{mm}$ ). While at highest dilution 1:75 all MDRPa isolates showed lowest activity of oil extract ( $\leq 10\text{ mm}$ ).

Disc containing absolute alcohol (diluent of the extract) produce no zone of inhibition. It was observed

that controlled strain was sensitive against black seed oil extract concentrations.

Aqueous extract of black seed at concentrations of 0.5% w/v, 0.05% w/v, 0.025% w/v and 0.005% w/v produced narrow diameter of inhibition zone (2-10 mm) which were not reproducible.

### Discussion:

*P. aeruginosa* remains the leading pathogen causing burn wound infection.<sup>21</sup> It survives well in the hospital environment. Once it is established, it can persist for months within a unit, posing as Multi drug resistant nosocomial infection risk for patients being treated there.<sup>22,23</sup> *Pseudomonas* is very resistant to most antibiotics and the resistance in this organism develops very rapidly. The rate of development of resistance to new antibiotics is much faster than the rate of invention and development of new antibiotics.<sup>24</sup>

Present study showed that the highest resistance was for Gentamicin 96.7%, this finding is similar to other studies conducted in Tohid burn centre Tehran Iran, and in Jinnah Postgraduate Medical Centre Karachi, where more than 95% and 93.2 strains of *Pseudomonas aeruginosa* were resistant to Gentamicin respectively.<sup>3, 21</sup> Gentamicin is a cheap and easily available drug that is used extensively in general and hospital practice in clinically suspected Gram-negative infections. This may be the main reason for the development of resistance in bacteria against this drug. Also the MDRPa isolates were highly resistant to the fourth generation Cephalosporins (Cefepime) 96.7% ,this result agree with Satti study which found that 71% of MDRPa isolated from various clinical specimens were resistant to Cefepime.<sup>25</sup> In the study of Mizuta *et al.*, *P. aeruginosa* isolates were most often susceptible to Ceftazidime (87% of isolates), Amikacin (84%),<sup>26</sup>which disagree partially to our results which showed high resistance to Ceftazidime (90%) but accepted for Amikacin (36.3%) resistance. Aztreonam is a monobactam  $\beta$ -lactam drug, it has excellent activity against *Pseudomonas* species but has a limited treatment option against MDR strains of *Pseudomonas aeruginosa* ,<sup>22</sup> the findings in the present study are in accordance with Douglas study, with about 83.3% resistance in MDRPa against this drug.<sup>22</sup> *P. aeruginosa* isolates showed resistance to Ciprofloxacin (63.3%), this might be due to the widespread prescribing of Fluoroquinolones in empirical therapy for *Pseudomonas* infections, which

may be associated with delays in administering effective therapy resulting in adverse outcomes. Similar results were noted in other studies.<sup>27</sup> Present study showed resistance to Levofloxacin (60% ), which agree with other study done by Kaye.<sup>28</sup> Piperacillin showed resistant ( 60%) to *Pseudomonas aeruginosa* isolates, this finding disagree to other study in which *Pseudomonas aeruginosa* remained 90% susceptible to Piperacillin.<sup>29</sup> Carbapenems (Imipenem, Meropenem) are useful in treatment of some cases of multi-drug resistant strains of *P. aeruginosa* ,<sup>22</sup> in this study the resistance of *P. aeruginosa* against Imipenem and Meropenem were 46.7% and 43.3% respectively. Similar results have been reported in other studies.<sup>1,17</sup> The highest sensitivity of *P. aeruginosa* was found to Colistin, this might be due to the less frequent use of this drug in the general practice because of the un sustained availability in hospitals and local markets.

In present study acetone extract yielded 20-30% of black seed oil .The ether extract made by Hanafy and Hatem,<sup>30</sup> was also dark brown and oily in nature but it was a clear liquid and yielded 11% v/w, while ether & methanolic extract made by Salman *et.al.*<sup>31</sup> yielded 16.7%, 32.5% respectively .These differences might be due to difference in the black seed sample, duration of soaking and / or temperature at which extraction was done.<sup>31</sup>

Our observation indicated that the antibacterial activity of oil extract against MDRPa isolates was dose dependent manner. This result is in accordance with earlier study.<sup>31</sup> The results of different concentration of black seed oil in present study showed direct relation with the diameter of inhibition zone and was found that acetone extract has clear effectiveness against MDRPa . However, all tested MDRPa isolates showed clear inhibitory effects of acetone extract at concentration 1:1, 1:10, 1:25 with inhibition zone of  $23.08 \pm 1.401$ ,  $19.482 \pm 2.014$ ,  $14.553 \pm 3.421$  respectively. Another study done by Salman tested oil and methanolic extract against *P. aeruginosa* isolated from different sources showed less significant effect at 1:1,1:10, the average size of inhibition zones were  $11 \pm 1$  and  $10 \pm 1$  respectively.<sup>11</sup> On contrary, the study was unable to achieve any degree of susceptibility to *P. aeruginosa* , *E.coli* and *Candida albicans* even with double or four times increase in volumes of oil or whole seed added.<sup>32</sup>This variation in antimicrobial activities might be due to that black

seed oil obtained from different commercial sources or isolated by different methods from the same seeds have been shown to vary significantly in their content of Thymoquinone, which has antibacterial activity and various storage conditions are expected to make a difference in the amounts of the quinone constituents of the oil, especially if the seed oil samples are exposed to heat and light.<sup>3,3,34</sup> Data presented in Table 3 indicated that 1:1,1:10,1:25 concentrations of oil extract resulted in highest antimicrobial activity produced zone of inhibition ranging from 16 - >25mm. Out of thirty MDRPa isolates tested with dilution 1:50 showed activity against nine isolates produce zone of inhibition 11-15mm. These results disagree with Salman results who mentioned that four and one strain inhibited by oil at concentration 1:1 produced 12-16mm and 17-21mm zone inhibition respectively.<sup>11</sup> Out of twenty one strains of *P. aeruginosa* tested , one was inhibited by oil at concentration 1:10 showed 12-16mm. The reason might be the differences in tested isolates. The antimicrobial effects of water extract showed the lowest and poor activity against all tested bacteria ,this result is in agreement with Salman *et al.*,<sup>31</sup> and it could be explained by the fact that the active ingredient, thymoquinone, is present in the oil extract, such an active substance may be missing or present in small amounts in the water extract .<sup>35</sup>

### **Conclusion and recommendation:**

The black seed oil extract using acetone is very effective as antibacterial agent against MDRPa and it gave better results than the oil extracted by other solvents. The water extract of black seed showed very little antibacterial activity against MDRPa. Although this study was done in vitro, the significant results obtained should direct us toward next step in using the oil extract in vivo, including topical application and / or systematic medication.

### **Knowledgments:**

I appreciate the cooperation of head & members of the Department of Microbiology / Faculty of Medical Sciences/Univ.of Duhok.I would like to express my thanks & full appreciation to Dr.Mahmoud ,Head Dep.of Chemistry / Faculty of Science/ Univ. of Duhok for providing laboratory facilities. I am deeply thankful to the staff of laboratory of Bacteriology in

Azadi Hospital and Burn& Plastic Surgery Hospital, for their assistance

#### References:

- 1-Song, W., Lee, K.M., Kang, H.J., Shin, D.H. and Kim, D.K. (2001). Microbiologic aspects of predominant bacteria isolated from the burn patients in Korea. *Burns*, 27(2): 136-139.
- 2-Barret, J.P., Gomex, P., Solano, I., Gonzalez-Derrego, M and Crisol, F.J. (1999). Epidemiology and mortality of adult burns in Catalonia. *Burns*, 25(4): 325-329.
- 3- Naqvi Z.A, Hashmi K., Rizwan Q.and Kharal S.A(2005).Multidrug resistant *Pseudomonas aeruginosa* :Anosocomial infection treat in burn patients. *Pakistan Journal of Pharmacology*.22(2) : 9-15.
- 4-Gang, R.K., Bang, R.L., Sanyal, S.C., Mokaddas, E. and Lari, A.R. (1999). *P. aeruginosa* septicaemia in burns. *Burns*, 25: 611-616.
- 5-Manson, W.L., Pernot, P.C.J., Fidler, V., Sauer, E.W. and Kalasen, H.J. (1992). Colonization of burns and the duration of hospital stay of severely burned patients. *J Hosp Infect*, 22: 55-63.
- 6-Baron, E. J., L. R. Peterson, and S. M. Finegold. (1994). Nonfermentative gram-negative bacilli and coccobacilli, p. 386-405. *Bailey & Scott's diagnostic microbiology*, 9th ed. Mosby-Year Book, Inc. St. Louis, MO.
- 7- Gilligan, P. H. (1995). *Pseudomonas* and *Burkholderia*, p. 509-519. In P. R. Murray, E. J. Baron, M. A. Pfaller, F. C. Tenover, and R. H. Tenover (eds.). *Manual of clinical microbiology*, 6th ed. American Society of Microbiology, Washington, D.C.
- 8- Vindenes, H. and Bjercknes, R. (1995). Microbial colonization of large wounds. *Burns*, 21(8): 575-579.
- 9-El-Shouny W.A. and Magnam S (2009).Sensitivity of multidrug resistant *Pseudomonas aeruginosa* isolated from surgical wound infections to essential oils and plant extracts.*World J.Med.Sci*,4(2),104-111.
- 10- Ali B,H, Blunden G. (2003). Pharmacological and toxicological properties of *Nigella sativa*. *Phytother Res*, 17:299–305.
- 11- Salman M.T, Khan R.A, Shukla I( 2008). Antimicrobial activity of *Nigella sativa* Linn seed oil against multi-drug resistant bacteria from clinical isolates. *Natural Product. Radiance*, 7(1): 10- 14.
- 12-Ara N, Choudhury S, Amin R ( 2005). In vitro Antimicrobial Activity of the Volatile Oil of *Nigella Sativa* Linn Seeds. *TAJ*, 18(2): 109-112
- 13-Arici M, Sagdic O,Gecgel U ( 2005). Antibacterial effect of Turkish black cumin (*Nigella sativa* L.) oils. *Grasas y Aceites*,56(4), 259-262.
- 14- National Committee for Clinical Laboratory Standards (NCCLS). ( 2002) Abbreviated identification of bacteria and yeast; Approved guideline. Wayne, PA: NCCLS; NCCLS document M35-A.
- 15-Brisse S.,Fluit A. C., Kusters K.,Klootwijk M.,De Vaal S.,Leverstein-van Hall M. A.,Verhoef J. and Milatovic D (2000). Evaluation of the Phoenix™ Automated Microbiology System for Detection of (i) Extended-Spectrum Beta-Lactamase-Producing *Escherichia coli* and *Klebsiella* spp., and of (ii) Ciprofloxacin-Resistant *Acinetobacter* spp. and *Pseudomonas aeruginosa* European Clinical Isolates.
- 16- Olayinka A.T, Onile B.A and Olayinka B.O (2004). Prevalence of multidrug-resistant *pseudomonas aeruginosa* isolates in surgical units of Ahmadu Bello University teaching hospital, Zaria,Nigeria: An indication for effective control measures. *Annals of African Medicine* 3(1) 13 – 16
- 17- Tam V.H, Chang K , Abdelraouf K, Brioso C.G, Ameka M, McCaskey LA, Weston JS, Caeiro J, Garey KW (2010). Prevalence, Resistance Mechanisms, and Susceptibility of Multidrug-Resistant Bloodstream Isolates of *Pseudomonas aeruginosa*. *Antimicrobial Agents And Chemotherapy*, 54( 3 )1160–1164.
- 18-Obritsch M.D, Fish D.N, MacLaren R and Jung R ( 2005).Nosocomial Infections Due to Multidrug-Resistant *Pseudomonas Aeruginosa*: Epidemiology and Treatment Options. *Pharmacotherapy*,25 (10) :1353-1364.
- 19- Ibraheem N K, Ahmed J H, Hassan M K ( 2010).The effect of fixed oil and water extracts of *Nigella sativa* on sickle cells: an in vitro study. *Singapore Med J*,51(3) : 230.
- 20- Pelczar M.J, Chan ECS, Krieg N.R (1999). *Microbiology Concepts and Applications*, 6th ed., McGraw-Hill Inc., New York, U.S.A.
- 21- Lari, A.K, Bahrami, H.H and Alaghebandan, R (1998). *Pseudomonas* infection in Tohid Burn Centre, Iran. *Burns*, 24(7):637-641.

22- Douglas,M.W, Mulholland K , Denyer V and Gottlieb T (2001). Multi-drug resistant *Pseudomonas aeruginosa* outbreak in a burns unit an infection control study. *Burns*, 27(2): 131-135.

23- Edwards V and Greenwood J. (2003). What's new in burn microbiology? James Laing Memorial Prize Essay. *Burns*,29(1): 15-24.

24- Estahbanati H.K, Kashani, P.P, and Ghanaatpisheh F. (2002). Frequency of *P.aeruginosa* serotypes in burn wound infections and their resistance antibiotics. *Burns*, 28(4): 340-348.

25- Satti L, Abbasi S, Kumar T.A, Khan M.S and Hashmi Z.A ( 2011).In Vitro Efficacy of Cefepime Against Multi-Drug Resistant *Pseudomonas aeruginosa*– an alarming situation in our setup. *The Open Drug Resistance Journal*, 1, 12-16.

26-Mizuta M, Linkin D.R, Nachamkin I, Fishman N.O, Weiner M.G, Sheridan A, Lautenbach E (2006). Identification of optimal combinations for empirical dual antimicrobial therapy of *P.aeruginosa* infection: potential role of a combination antibiogram. *Infect Control Hosp Epidemiol* 27(4):413-5.

27-Hsu DI,Okamoto M.P, Murthy R, Wong-Beringer A. ( 2005). Fluoroquinolone-resistant *pseudomonas aeruginosa*: Risk factors for acquisition and impact on outcomes. *J Antimicrob Chemother*,55(4):535-41.

28-Kaye KS, Kanafani Z.A, Dodds AE, Engemann J.J, Weber S.G, Carmeli Y (2006 ) .Differential effects of levofloxacin and ciprofloxacin on the risk for isolation of quinolone-resistant *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother*, 50(6):2192-6.

29- Walton, M.A., Villarreal, C., Herndon, D.N. and Hegggers, J.P. (1997). The use of aztreonam as an alternative therapy for multi-resistant *P. aeruginosa*. *Burns*, 23(3): 225-7.

30- Hanafy M.S.M and Hatem M.E (1991) .Studies on the antimicrobial activity of *Nigella sativa* seed (Black Cumin). *J Ethnopharmacol*, 34(2-3), 275-278.

31-Salman M.T, Khan R.A, Shukla I. A ( 2009) . Study of *Nigella sativa* Linn. seeds for antimicrobial activity against multidrug resistant clinical strains of *Pseudomonas aeruginosa*. *Hippocratic Journal of Unani Medicine*. 4(4) 95-104.

32- Salman A.A, Hosny A.M.S, Ahmady AM, Abedl-Hamid AA, Hussien ASM( 2004) Suceptibility of nosocomial pathogens to certain antibiotics and biocides commonly used in Egyptian hospitals. *N.Egypt.J.Microbiol*. 8,274-299.

33-Aboul E.H.Y and Abou B.L.I (1995). Simple HPLC method for the determination of thymoquinone in Black Seed oil (*Nigella sativa* Linn). *J Liquid Chromatography*, 18(5) 895-902.

34-Abou B.L.I, Rashed MS and Aboul EHY, TLC (1995). Assay of thymoquinone in Black Seed oil (*Nigella sativa* Linn) and identification of dithymoquinone and thymol. *J Liquid Chromatography*,18(1),105-115.

35- Ghosheh O.A, Houdi A.A, Crooks P.A.( 1999 ). High performance liquid chromatographic analysis of the pharmacologically active quinones and related compounds in the oil of the black seeds( *nigella sativa* L.). *J Pharm Biomed Anal* ,19:757-62.

Instrument ID(s)	Confidence Value	Biochemical		Expected Result		Biochemical		Expected Result	
Pseud. aeruginosa	99%	Instr. Result	Expected Result	Instr. Result	Expected Result	Instr. Result	Expected Result	Instr. Result	Expected Result
A_ABARR	+	+	V	A_GUPBB	-	A_GURV	-	A_GURV	-
A_GNDAA	-	-	-	A_LABGE	+	A_LGFA	+	A_LGFA	+
A_LERHH	-	-	-	A_LPHRE	-	A_LPHRO	-	A_LPHRO	-
A_LPFR	+	+	V	A_LTBV	-	A_LVALD	-	A_LVALD	-
C_AACT	-	-	-	C_AAO	-	C_COT	-	C_COT	-
C_C3CF	-	-	-	C_DAMT	-	C_RGA	-	C_RGA	-
C_ABEU	-	-	-	C_PBB	-	C_TBT	-	C_TBT	-
M_NAG	-	-	-	N_LGGH	+	N_LPROT	+	N_LPROT	+
R_BREAU	-	-	-	R_BPHS	-	R_BBLU	-	R_BBLU	-
R_BREU	-	-	-	R_DIN	-	R_DIBU	-	R_DIBU	-
R_BREU	-	-	-	R_DJHA	-	R_DJBL	-	R_DJBL	-
R_DSMF	-	-	-	R_DMAC	-	R_ORA	-	R_ORA	-
R_ARD	-	-	-	R_2HHA	-	R_SMBL	-	R_SMBL	-
R_MFU	-	-	-	R_NHA	-	R_NGU	-	R_NGU	-
S_ORN	-	-	-	S_URE	-	T_DSC	-	T_DSC	-

**Table .1: Identification of *P. aeruginosa* by BD - PhoenixTM Automated Microbiology System (original report).**

Antimicrobial	Instr. MIC	Instr. SIR	BDxprt SIR	Final SIR	Risk #
Ambicid	>32	R	R	R	
Carbamocin	>32	R	R	R	
Etoposim	>4	R	R	R	
Imipenem	>8	R	R	R	
Margiprom	>8	R	R	R	
Ceftazidim	>16	R	R	R	
Ceftazidim	>16	R	R	R	
Ceftazidim	>32	R	R	R	
Cefepime	>16	R	R	R	
Cefepime-Sulbactam	>328	R	R	R	
Aztreonam	>16	R	R	R	
Amoxicillin-Sulbactam	>1608	R	R	R	
Piperacillin-Tazobactam	644	R	R	R	
Ceftriaxone	<=1	S	S	S	
Trimethoprim-Sulfamethoxazole	>4/76	R	R	R	
Ciprofloxacin	>2	R	R	R	
Levofloxacin	>4	R	R	R	
Tetracycline	>8	R	R	R	

**Table .2: Antibiotic sensitivity of *P. aeruginosa* by BD - Phoenix™ Automated Microbiology System (original report).**



**Fig.3:** Revealed to susceptibility of *P. aeruginosa* to different concentrations of black seed oil impregnated filter paper on MuellerHinton Agar Plate.

04	08	05	04	04	05
(13.3%)	(26.6%)	(16.6%)	(13.3%)	(13.3%)	(16.6%)

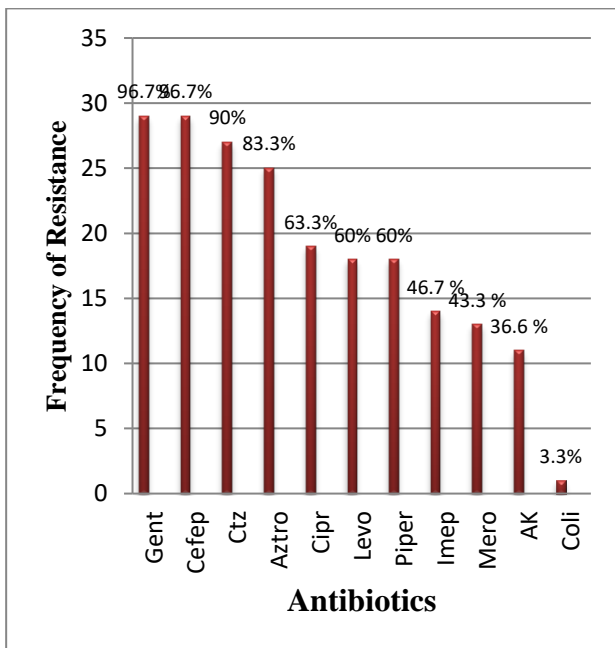
**Table 2:** Inhibition zones (measured as mm) of different concentrations of black seed oil extract against MDRPa.

n = 30	Black seed oil extract (zones in mm)				
	MDRPa				
	1:1	1:10	1:25	1:50	1:175
Mean ± SEM	23.08* ± 1.401	19.482* ± 2.014	14.553* ± 3.421	7.882 ± 4.818	4.077 ± 3.471
Mean ± SEM					
Mean ± SEM					
Mean ± SEM					
Mean ± SEM					

\* P < 0.05

**Table 3:** Sensitivity pattern of MDRPa to different concentrations of black seed oil extract

Total No. of isolates	Zone of inhibition (mm)	Black seed oil concentrations				
		1:1	1:10	1:25	1:50	1:75
30	>25	7	1	-	-	-
30	21-25	19	11	-	-	-
30	16-20	4	16	17	-	-
30	11-15	-	2	6	9	-
30	≤ 10	-	-	7	21	30



**Fig. 4 :** Resistant pattern of *Pseudomonas aeruginosa* isolates .

( Gent- Gentamicin , Cefep- Cefepime ,Ctz- Ceftazidime , Aztro- Aztreonam, Cipr-Ciprofloxacin ,Levo- Levofloxacin , Piper- Piperacilli, Imep-Imipenem , Mero- Meropenem ,AK- Amikacin , Coli- Colistin).

**Table 1:** Distribution of resistance among 30 *P. aeruginosa* isolates against 11 antimicrobial drugs.

Resistant to 4	Resistant to 5	Resistant to 6	Resistant to 8	Resistant to 9	Resistant to 10
----------------	----------------	----------------	----------------	----------------	-----------------

## تأثير الفعالية المقاومة للميكروبات لمستخلص الحبة السوداء المائي والزيتي على بكتيريا الزائفة الزنجارية المتعددة المقاومة للمضادات

منى سلمان الدليمي

### الخلاصة

تعد جرثومة الزائفة الزنجارية احدى عدوى المستشفيات الاكثر شيوعا في مراكز الحروق . وتشكل هذه الجرثومة احدى المخاطر التي تهدد حياة مرضى الحروق اذ كونها انتهازية ومعروفة لمقاومتها المضادات الحيوية. جمعت عينات الدراسة للمرضى الراقدين في مستشفى الحروق والجراحة التجميلية في دهوك\العراق ,لفترة من ايلول 2011 ولغاية اذار 2012، وتم استخدام الطرائق التقليدية للتشخيص التمهيدي لهذه الجرثومة وعززت باستخدام جهاز BD – PhoenixTM، واعتبرت هذه العزلات من الجراثيم متعددة المقاومة للمضادات ,اذا اظهرت مقاومة لثلاث اواكثر من المضادات التي تستخدم في علاج الخمج الناجم عن الاصابة بهذه الجراثيم ووفقا لذلك عزلت ثلاثون عزلة متعددة المقاومة للمضادات خلال الدراسة .واظهرت العزلات اعلى نسبة مقاومة للمضادات جنتاميسين وسيفيبيم و سيفتازديم وازترونام (96.7%، 96.7% و 90% و 83.3%) على التوالي وكان المضاد اميكسين وكوليستين الاقل مقاومة (36.7% 3.3%). وقد اختبرت الفعالية المضادة للجراثيم لمستخلص الحبة السوداء المائي والزيتي على جراثيم الزائفة الزنجارية المتعددة المقاومة للمضادات .واظهر زيت الحبة السوداء تركيز 1:1, 1:10, 1:25 ,فعالية مضادة للجراثيم اكثر معنوية من زيت الحبة السوداء بتركيز 1:75 وكانت الفعالية المضادة للجراثيم للمستخلص المائي منخفضة وقليلة على جميع العزلات المدروسة.